Tetrahedron 58 (2002) 3619-3622

Diterpenylated and prenylated flavonoids from Macaranga denticulata

Somyote Sutthivaiyakit, a,* Sasinee Unganont, Pakawadee Sutthivaiyakit and Apichart Suksamrarn

^aDepartment of Chemistry, Faculty of Science, Ramkhamhaeng University, Huamark, Ramkhamhaeng Road, Bangkapi, Bangkok 10240, Thailand

^bDepartment of Chemistry, Faculty of Science, Kasetsart University, Bangkok 10900, Thailand

Received 27 November 2001; revised 6 February 2002; accepted 7 March 2002

Abstract—An *O*-methylated analogue of macarangin (1) and denticulaflavonol (2) which possesses a diterpene substituent have been isolated along with other compounds from the leaves of *Macaranga denticulata*. Their detailed 1D and 2D spectral analysis are reported. The radical scavenging properties of compounds 3–5 were evaluated. Among the compounds tested, 4 showed a pronounced antioxidant activity (IC $_{50}$ 0.032 \pm 0.001 mM). © 2002 Elsevier Science Ltd. All rights reserved.

In our investigation for bioactive constituents from plants, we studied the leaves of *Macaranga denticulata* Muell. Arg. (synonymous name *Macaranga henricorum* Hemsl., Euphorbiaceae), known in Thai as 'Tong Taeb'. The plant is a shrubby tree which grows to about 20 m in height. The stem water decoction has been used traditionally for washing wounds and drunk as tonic by women after child labor. Previously reported constituents from the species of this genus comprise terpenes, steroids, hydrolysable tannins and prenylflavanones from *M. tanarius*, bergenin derivatives from *M. peltata*, chromenoflavones from *M. indica*, a hexahydroxanthene derivative and a geranyl flavonol from *M. vedeliana*, prenylated flavanones from *M. pleiostemona*, and geranyl stilbenes from *M. schweinfurthii*.

Two flavonoids, one of which is named denticulaflavonol, features the presence of a unique labdane diterpene moiety, have been isolated from the leaves of this plant along with other compounds. To our knowledge, this is the first example of a flavonoid possessing a diterpene substituent. We herein describe the isolation, structural elucidation and radical scavenging properties of the isolates.

The chloroform extract of the dried leaves of M. denticulata was purified as described in the experimental section to yield two compounds (1, 2) as well as scopoletin (3), macarangin (4)⁸ and sophoraflavanone B (5). The known

compounds were identified by comparison of their published spectral data.

Compound 1 was obtained as yellow sticky solid. It was assigned the molecular formula $C_{26}H_{28}O_6$ by HR-EIMS. The FT-IR showed absorption bands at $\nu_{\rm max}$ 3275, 1658 and 1608 cm⁻¹ indicating the presence of a hydroxy, an

Keywords: Macaranga denticulata; Euphorbiaceae; diterpene; flavonoid; diterpenylated flavonoid; antioxidant.

^{*} Corresponding author. Tel.: +662-319-1900; fax: +662-310-8401; e-mail: somyote_s@yahoo.com; somyote@ram1.ru.ac.th

Table 1. ¹H and ¹³C NMR spectral data of **1** (in MeOH-d₄), **2** and **4** (in CDCl₃)

| No. | 2 | | | 1 | | | 4 | |
|-------|------------------------------|--------------------------|------------------------------|--------------------------------|--------------------------|------------------|------------------------------|--------------------------|
| | $\delta_{ m H}{}^{ m a}$ | ${\delta_{ m C}}^{ m b}$ | НМВС | $\delta_{	ext{H}}{}^{	ext{a}}$ | ${\delta_{ m C}}^{ m b}$ | HMBC | $\delta_{	ext{H}}^{	ext{a}}$ | ${\delta_{ m C}}^{ m b}$ |
| 2 | | 146.0 (s) | | | 130.6 (s) | | | 145.5 (s) |
| 3 | 6.59 (s, 3-OH) | 135.5 (s) | 3-OH/C-2, 3, 4 | 3.66 (s, 3-OMe) | $139.8 (s)^{c}$ | 3-OMe/C-3 | 5.9 (s, 3-OH) | 135.4 (s) |
| 4 | | 175.3 (s) | | | 179.9 (s) | | | 175.2 (s) |
| 5 | 12.1 (s, 5-OH) | 158.0 (s) | 5-OH/C-5, 6, 10 | | 159.7 (s) | | 12.1 (s, 5-OH) | 157.4 (s) |
| 6 | | 109.4 (s) | | | 112.9 (s) | | | 109.4 (s) |
| 7 | 6.25 (s, 7-OH) | 161.7 (s) | 7-OH/C-6, 7, 8 | | 163.7 (s) | | 6.6 (s, 7-OH) | 161.8 (s) |
| 8 | 6.48 (s) | 94.3 (d) | C-6, 9, 10 | 6.33 (s) | 93.9 (d) | C-4, 6, 7, 9, 10 | 6.45 (s) | 94.3 (d) |
| 9 | | 155.0 (s) | | . , | 156.3 (s) | | ` ' | 155.0 (s) |
| 10 | | 103.2 (s) | | | 105.7 (s) | | | 103.5 (s) |
| 1' | | 123.5 (s) | | | 122.7 (s) | | | 123.4 (s) |
| 2',6' | 8.12 (d, 8.9) | 129.6 (d) | C-2, 4' | 7.87 (d, 8.9) | 131.3 (d) | C-2, 3', 5' | 8.1 (d, 8.5) | 129.6 (d) |
| 3',5' | 6.97 (d, 8.8) | 115.6 (d) | C-1', 4' | 6.82 (d, 8.9) | 116.5 (d) | C-2', 4' | 6.94 (d, 8.3) | 115.6 (d) |
| 4′ | 5.37 (s, 4'-OH) | 157.3 (s) | 4'-OH/C-3', 5' | , , , | 161.6 (s) | | 5.5 (s, 4'-OH) | 157.6 (s) |
| 1" | a 1.74 (m), b 0.97 (ddd, | 39.0 (t) | C-3", 5", 9", 20" | 3.22 (m) | 22.2 (t) | C-5, 7, 2", 3" | 3.48 (d, 7.1) | 21.4 (t) |
| | 12.8, 12.8, 3.9) | | | | | | , , , | |
| 2" | n.o. | n.o. | | 5.14 (dd, 7.2, 6.3) | 123.5 (d) | C-6, 4", 5" | 5.28 (m) | 120.9 (d) |
| 3" | a 1.37 m, b 1.15 (ddd, | 42.1 (t) | C-1", 18", 19" | , , , , , | 135.7 (s) | | ` ' | 139.8 (s) |
| | 13.4, 13.2, 4.0) | | | | | | | |
| 4" | | n.o. | | 1.68 (s) | 16.3 (q) | C-3", 5" | 1.83 (s) | 16.2 (q) |
| 5" | 1.05 (dd, 9.9, 2.7) | 55.5 (d) | C-1", 6", 10", 18", 19", 20" | 1.86 (dd, 7.4, 6.8) | 40.9 (t) | C-2", 3", 4", 6" | 2.10 (m) | 39.7 (t) |
| 6" | a 1.67 (m), b 1.27 (m) | 24.4 (t) | C-8" | 1.95 (dd, 7.6, 7.1) | 27.7 (t) | C-3", 5", 7" | 2.10 (m) | 26.3 (t) |
| 7" | a 2.4 (ddd, 12.7, 4.1, 2.3), | 38.3 (t) | C-5", 6", 8", 17" | 4.95 (m) | 125.4 (d) | C-9", 10" | 5.05 (m) | 123.7 (d) |
| | b 1.90 (ddd, 13.7, 13.3, | | | | | | ` ' | |
| | 4.7) | | | | | | | |
| 8" | ŕ | 148.6 (s) | | | 132.0 (s) | | | 132.1 (s) |
| 9" | 1.56 (m) | 56.0 (d) | C-1", 8", 17", 20" | 1.51 (s) | 25.8 (q) | C-7", 10" | 1.68 (s) | 25.7 (q) |
| 10" | . , | 39.6 (s) | | 1.45 (s) | 17.7 (q) | C-7", 9" | 1.59 (s) | 17.7 (q) |
| 11" | a 1.47 (m), b 1.43 (m) | 19.4 (t) | C-9", 12" | . , | | | ` ' | |
| 12" | a 2.19 (m), b 1.85 (m) | 38.5 (t) | C-13" | | | | | |
| 13" | | 140.7 (s) | | | | | | |
| 14" | 5.27 (dd, 7.3, 6.5) | 120.6 (d) | C-12", 16" | | | | | |
| 15" | 3.49 (d, 7.0) | 21.4 (t) | C-5, 6, 7, 13", 14" | | | | | |
| 16" | 1.84 (s) | 16.4 (q) | C-12", 13" | | | | | |
| 17" | a 4.80 (d, 1.2), b 4.49 (s) | 106.2 (t) | C-7", 8" | | | | | |
| 18" | 0.86 (s) | 33.6 (q) | C-3", 19" | | | | | |
| 19" | 0.79 (s) | 21.7 (q) | C-3", 18" | | | | | |
| 20" | 0.67 (s) | 14.5 (q) | C-1", 5", 9" | | | | | |
| | | (-1) | - ,- ,- | | | | | |

n.o. signal was not observed, may be obscured by other signals.

α,β-unsaturated carbonyl and aromatic groups, respectively. The ¹H NMR in MeOH-d₄ exhibiting one set of AB doublets at $\delta_{\rm H}$ 7.87 (2H, d, J=8.9 Hz) and 6.82 (2H, d, J=8.9 Hz) together with the ¹³C NMR signals at $\delta_{\rm C}$ 161.6 (s), 131.3 (d), 116.5 (d) and 122.7 (s) indicated the paradisubstituted phenol ring. The presence of a geranyl group was deduced from the mass spectrum which showed base peak at m/z 313 ([M-CH=CMeCH₂CH₂CH=C(Me)₂]⁺, 100 %) as well as the ¹H NMR showing two olefinic protons at $\delta_{\rm H}$ 5.14 (1H, d, J=7.2, 6.3 Hz) and 4.95 (1H, m), allylic protons at $\delta_{\rm H}$ 3.22 (2H, d, J=3.4, 1.7 Hz), two sets of two proton signals at $\delta_{\rm H}$ 1.95 (dd, J=7.6, 7.1 Hz) and 1.86 (dd, J=7.4, 6.8 Hz) and three methyl group signals at $\delta_{\rm H}$ 1.68 (s), 1.51 (s), and 1.45 (s). The lowfield signal at δ_H 12.98, an indicative evidence of a C-5-OH proton intramolecularly H-bonded to C-4 carbonyl oxygen atom of a flavonoid nucleus, could be detected when deuterated chloroform was used as solvent although the compound is sparingly soluble. A broad signal at $\delta_{\rm H}$ 6.33 assigned to the C-8 H showed ${}^{3}J_{\text{CH}}$ correlations to the signals at δ_{C} 105.7 (C-10), 112.9 (C-6) as well as ${}^2J_{\text{CH}}$ and ${}^4J_{\text{CH}}$ correlations to the signals at δ_C 156.3 (C-9), 163.7 (C-7) and 179.9 (C-4), respectively. A geranyl substitution at C-6 was deduced from the important $^3J_{\text{CH}}$ correlations of the signal at δ_{H} 3.22 (H-1") to ^{13}C resonances at δ_{C} 112.9 (C-6), 123.5 (C-2"), 135.7 (C-3") 159.7 (C-5) and 163.7 (C-7). This compound showed NMR data very similar to values published for macarangin (4), a compound previously isolated from M. $vedeliana^8$ and also isolated in the present study. The only difference observed was a methyl proton signal at δ_{H} 3.66. The $^3J_{\text{CH}}$ correlation between the signal at δ_{H} 3.66 with the signal at δ_{C} 139.8 (C-3) strongly indicated the presence of an O-methyl group at C-3. Compound 1 was proposed to be 3-O-methyl-macarangin, 3-O-methyl-6-[(E)-3",8" dimethyl-2",7"-octadienyl]-kaempferol.

Compound **2** was obtained as yellowish sticky mass. The HR-EIMS gave a molecular formula of $C_{35}H_{42}O_6$. The IR spectrum showed the presence of a hydroxyl and a carbonyl groups at $\nu_{\rm max}$ 3232 and 1647 cm⁻¹, respectively. The ¹H NMR spectrum measured in CDCl₃ showing two sets of doublets at $\delta_{\rm H}$ 8.12 (H-2', 6') and 6.97 (H-3', 5'), a one proton singlet at $\delta_{\rm H}$ 6.48 (H-8) and a downfield proton at $\delta_{\rm H}$ 12.1 (C-5–OH) with the absence of the 3-OMe group

^a Multiplicities and *J*-values in Hz are given in parentheses.

b Multiplicities were deduced from DEPT experiments.

^c Signal of 3-OCH₃ was found at $\delta_{\rm C}$ 60.5 (q).

signal, as found in compound 1, indicated the existence of a kaempferol C₁₅H₉O₆ unit with the same substitution pattern as for 1. The remaining $C_{20}H_{31}$ moiety, establishing five double bond equivalents, was recognized as a labdane type diterpene conjugate by the presence of an exocyclic methylene group signals at $\delta_{\rm H}$ 4.80 (1H, d, J=1.2 Hz) and 4.49 (1H, s), a vinylic proton of a trisubstituted double bond at $\delta_{\rm H}$ 5.27 and a downfield methyl group signal at $\delta_{\rm H}$ 1.84 (3H, s) assignable to $-C=C-CH_3$ in addition to the three shielded quaternary methyl group singlets at $\delta_{\rm H}$ 0.86, 0.79 and 0.67 which have one-bond correlations with the ¹³C shifts of the methyl groups commonly encountered in the labdanic diterpenes 14 at δ_C 33.6 (C-18″), 21.7 (C-19″) and 14.5 (C-20") in the HMQC spectrum, respectively. Connection of the labdanyl unit to C-6 was established by the long range ${}^{1}H$ – ${}^{13}C$ correlations of signals at $\delta_{\rm H}$ 3.49 (assigned for H-15") to signals at $\delta_{\rm C}$ 109.4 (C-6), 158.0 (C-5) and 161.7 (C-7). The placement of a trisubstituted double bond at C-13"-C-14" was further deduced by the ¹H-¹H COSY and HMBC correlations particularly of H-15" to C-13" $(\delta_{\rm C}\ 140.7)$ and C-14" $(\delta_{\rm C}\ 120.6)$. Detailed interpretation of the ¹H-¹H COSY, HMQC and HMBC spectra, in conjunction with the analysis of ¹³C shifts of several representatives of the labdane diterpenes reported previously, 14 permitted unambiguous assignments of ¹H and ¹³C resonances as indicated in Table 1, the key long range $^{1}H^{-13}C$ correlations are also shown. Compound 2, trivially named denticulaflavonol, was identified as 6-[15"-labda-8" (17"),13" (14")-dienyl]-kaempferol.

Due to the scarcity of pure samples, only compounds 3-5 were evaluated for their antioxidant properties using 1,1-diphenyl-2-picrylhydrazyl (DPPH) stable radical. Macarangin (4) exhibited an IC₅₀ 0.032 \pm 0.001 mM which is as pronounced as a standard antioxidant, 2,6-di-(*tert*-butyl)-4-methylphenol (BHT), IC₅₀ 0.031 \pm 0.001 mM. Scopoletin (3) also showed rather significant antioxidant property (IC₅₀ 0.342 \pm 0.026 mM). Since sophoraflavanone B (5) showed only 31.8% reduction at 0.759 mM, its IC₅₀ was therefore not determined.

1. Experimental

1.1. General

The specific rotations were measured by a Jasco DIP 1020 polarimeter. The IR spectra were obtained on a Perkin–Elmer 1760x FT-IR spectrophotometer. UV spectra were recorded on a Perkin–Elmer Lambda 6 instrument. EI-MS and HR-EIMS spectra were recorded on a Finnigan MAT 90 instrument. ¹H and ¹³C spectra were obtained with a Bruker AVANCE 400 MHz spectrometer with the solvent signal as internal reference.

1.2. Plant material

Leaves of *M. denticulata* were collected from Trat Province in December 1997. The species was kindly identified by Dr K. Chayamarit of the Royal Forest Herbarium, Bangkok. A voucher specimen (SSMD/1997) is kept at the Chemistry Department, Faculty of Science, Ramkhamhaeng University.

1.3. Extraction and isolation

The dried leaves (1230 g) were ground and extracted successively with *n*-hexane, chloroform and methanol, respectively. The chloroform extract (33.7 g) was separated on silica gel with gradient of n-hexane/CH₂Cl₂ (15:85) to CH₂Cl₂/MeOH (3:2) to afford nine fractions. Fraction 8 was purified by column chromatography over a Si gel, with gradient of *n*-hexane/EtOAc (3:1) to EtOAc/MeOH (9:1) to obtain six fractions (fractions 8.1-8.6). Additional Si gel chromatography of fraction 8.3 with n-hexane/EtOAc (9:1) afforded 12 subfractions (subfractions 8.3.1–8.3.12). Compounds 1 (20 mg, 0.00163% w/w) and 5 as a sticky gum (31 mg, 0.00252% w/w) were obtained from subfraction 8.3.6 after further column chromatography over Si gel (n-hexane/EtOAc 8:2). Fraction 6 was chromatographed over Si gel, eluted with a gradient of CH₂Cl₂ to CH₂Cl₂/MeOH (6:4) and afforded subfractions 6.1-6.5. Subfraction 6.4 was further chromatographed over Si gel using gradient of n-hexane/CH₂Cl₂ 2:8 to CH₂Cl₂ and yielded subfractions 6.4.1-6.4.3. Subfraction 6.4.1 contained pure compound 3 as a pale yellow solid, mp 193–195°C, (20 mg, 0.00163% w/w). Subfraction 6.4.2 gave compound 2 (3 mg, 0.00024% w/w) after additional column chromatography over Si gel using *n*-hexane/EtOAc 9:1. Subfraction 6.4.3 was purified by Si gel column chromatography using gradient of n-hexane/CH₂Cl₂ 2:8 to CH₂Cl₂/MeOH 9:1 to yield compound 4 as a yellow solid, mp 140-143°C, (120 mg, 0.00975% w/w).

1.3.1. 3-*O*-Methyl-macarangin (1). Yellow sticky solid; $R_{\rm f}$ 0.17 [silica gel, n-hexane/EtOAc 7:3]; $[\alpha]_{\rm D}^{26} = -0.5254$ (c 0.590, MeOH); UV (MeOH) $\lambda_{\rm max}$ (log ε): 204 (4.50), 214 (4.49), 270 (4.27), 339 (4.23) nm; IR (film) $\nu_{\rm max}$: 3275, 2936, 1658, 1608, 1568, 1471, 1359, 1286, 1233, 1177, 1089, 973, 815 cm⁻¹; ¹H and ¹³C NMR data (measured in MeOH- d_4) see Table 1; EI-MS (70 eV) m/z (rel. int.): 437 ([M+1]⁺, 1) 371 (21), 365 (11), 351 (18), 345 (21), 344 (61), 340 (16), 325 (15), 314 (20), 313 (100), 299 (10), 281 (5), 270 (13), 253 (6), 242 (9), 203 (10), 165 (8), 121 (55), 105 (11), 91 (18); HR-EIMS m/z 436.1894 [M]⁺ (calcd for $C_{26}H_{28}O_6$, 436.1886, error in ppm=1.83).

1.3.2. Denticulaflavonol (2). Yellow sticky mass; $R_{\rm f}$ 0.32 [silica gel, n-hexane/EtOAc 7:3]; $\left[\alpha\right]_{\rm D}^{26} = -131.8$ (c 0.050, CHCl₃); UV (MeOH) $\lambda_{\rm max}$ (log ε) 207 (4.67), 270 (4.13), 329 (4.04) nm; IR (film) $\nu_{\rm max}$: 3232, 2921, 1647, 1542, 1508, 1363, 1180, 1087, 883, 841 cm⁻¹; ¹H and ¹³C NMR data (measured in CDCl₃) see Table 1; HR-EIMS m/z 558.2978 [M]⁺ (calcd for C₃₅H₄₂O₆, 558.2981, error in ppm=0.53).

1.4. Bioassay

Compounds 3–5 were tested for radical scavenging properties using DPPH. 15 50 μ L of a solution containing the compound to be tested was added to 5 ml of a 0.004% methanolic solution of DPPH. Absorbance at 517 nm was determined after 30 min at 37°C, and the percent of activity was calculated. IC₅₀ is the mean \pm standard deviation of three assays.

Acknowledgements

We are grateful to the Thailand Toray Science Foundation, the Thailand Research Fund and Ramkhamhaeng University for financial support. Thanks are due to Dr K. Chayamarit of the Forest Herbarium, Royal Forest Department, Bangkok, for the identification of the plant material, Mr N. Chimnoi, Chulabhorn Research Institute, for HRMS measurements and to Ms P. Srisomphot for some technical assistance.

References

- 1. Phupattanapong, L.; Wongprasert, T. *Thai Medicinal Plants*. *Part 5.*; Chutima: Bangkok, 1987, p 687 (in Thai).
- Hui, W. H.; Ng, K. K.; Fukamiya, N.; Koreeda, M.; Nakanishi, K. Phytochemistry 1971, 10, 1617–1620.
- Hui, W. H.; Li, M. M.; Ng, K. K. Phytochemistry 1975, 14, 816–817.
- Lin, J. H.; Nonaka, G. I.; Nishioka, I. Chem. Pharm. Bull. 1990, 38, 1218–1223.
- Tseng, M. H.; Chou, C. H.; Chen, Y. M.; Kuo, Y. H. J. Nat. Prod. 2001, 64, 827–828.

- Ramaiah, P. A.; Row, L. R.; Reddy, D. S.; Anjaneyulu, A. S. R. J. Chem. Soc., Perkin Trans. 1 1979, 2313–2316.
- 7. Sultana, S.; Ilyas, M. Phytochemistry 1986, 25, 953-954.
- 8. Hnawia, E.; Thoison, O.; Gueritte-Voegelein, F.; Bourret, D.; Sevenet, T. *Phytochemistry* **1990**, *29*, 2367–2368.
- 9. Thoison, O.; Hnawia, E.; Gueritte-Voegelein, F.; Sevenet, T. *Phytochemistry* **1992**, *31*, 1439–1442.
- Schutz, B. A.; Wright, A. D.; Rali, T.; Sticher, O. *Phytochemistry* 1995, 40, 1273–1277.
- Beutler, J. A.; Shoemaker, R. H.; Johnson, T.; Boyd, M. R. J. Nat. Prod. 1998, 61, 1509–1512.
- Pouchert, C. J.; Behnke, J. 1st ed.; The Aldrich Library of ¹³C and ¹H FT NMR Spectra, Vol. 2; Aldrich Chemical, Wisconsin, 1993 p 1314.
- Komatsu, M.; Yokoe, I.; Shirataki, Y. Chem. Pharm. Bull. 1978, 26, 3863–3870.
- Buckwalter, B. L.; Burfitt, I. R.; Nagel, A. A.; Wenkert, E.;
 Naf, F. Helv. Chim. Acta 1975, 58, 1567–1573.
- Atta-Ur-Rahman; Ngounou, F. N.; Choudhary, M. I.; Malik, S.; Makhmoor, T.; Nur-E-Alam, M.; Zareen, S.; Lontsi, D.; Ayafor, J. F.; Sondengam, B. L. *Planta Med.* 2001, 67, 335–339.